



Mini Review

PrimPol (Primase/Polymerase), replicating enzyme – A mini review

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Abstract

Polymerases were revealed first in 1970s. Most important to the modest perception the enzyme responsible for nuclear DNA replication that was pol α , for DNA repair pol β and for mitochondrial DNA replication pol γ . DNA construction and renovation done by DNA polymerases, so directing both the constancy and discrepancy of genetic information. Replication of genome initiate with DNA template-dependent fusion of small primers of RNA. This preliminary phase in replication of DNA demarcated as *de novo* primer synthesis which is catalyzed by specified polymerases known as primases. Sixteen diverse DNA-synthesizing enzymes about human perspective are devoted to replication, reparation, mutilation lenience, and inconsistency of nuclear DNA. But in dissimilarity, merely one DNA polymerase has been called in mitochondria. It has been suggest that PrimPol is extremely acting the roles by re-priming DNA replication in mitochondria to permit an effective and appropriate way replication to be accomplished. Investigations from a numeral of test site have significantly amplified our appreciative of the role, recruitment and regulation of the enzyme during DNA replication. Though, we are simply just start to increase in value the versatile roles that play PrimPol in eukaryote.

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INTRODUCTION

Multifaceted cellular tasks are executed by complexes of protein technologies. These protein associations comprehend vastly in step moving parts. In general whose tasks are temporally and spatially controlled by successions of systematic conformational vagaries and resulting from hydrolysis of nucleoside triphosphates are driven by chemical energy (1, 2). Polymerases (pols β - γ) were revealed first in 1970s. Most important to the modest perception the enzyme responsible for nuclear DNA replication that was pol α , for DNA repair pol β and for mitochondrial DNA replication pol γ (3). DNA construction and renovation done by DNA polymerases, so directing both the constancy and discrepancy of genetic information (4, 5). Replication of genome initiate with DNA template-dependent fusion of small primers of RNA using. This preliminary phase in replication of DNA demarcated as *de novo* primer synthesis which is catalyzed by specified polymerases known as primases. Sixteen diverse DNA-synthesizing enzymes about human perspective are devoted to replication, reparation, mutilation lenience, and inconsistency of nuclear DNA. But in dissimilarity, merely one DNA polymerase has been called in mitochondria (2, 6). Each DNA synthesis reaction associated to the replication and reparation of mitochondrial DNA (mtDNA), DNA polymerase gamma (Poly γ), which has long been supposed to intervene (5).

An Archaic Primase/Polymerase (PrimPol)

PrimPol (primase/polymerase) the additional primase to be discovered in human cells and is existing in both nuclear and

mitochondrial sections (5). PrimPol as name suggests can task equally as a primase and a polymerase. Various revisions have verified that PrimPol is proficient of making both DNA and RNA primers on single-stranded (ss) DNA templates (7). Up to newly, Pol α -associated DNA primase small subunit (PriS) liable for *de novo* polymer synthesis over the construction of RNA primers was reflected to be the only AEP (archaeo-eukaryotic primase) superfamily associate existing in eukaryotes (8). Evolutionarily distinct relations primases can be distributed in two: DnaG-like (prokaryotic primases) and AEP-like (archaeo-eukaryotic primases) (9). Though, bioinformatics investigation acknowledged further actuality of un-described DNA primase in eukaryotes known as PrimPol (CCDC111 or FLJ33167) which fit in to 'NCLDV-herpes virus clade' of viral AEPs (archaeo-eukaryotic primases) (5, 10-13).

PrimPol (Primase/Polymerase) and Lesion Bypass Venture PrimPol Act as a Primase

In silico findings prophesied primarily that for primer synthesis, PrimPol is a dynamic primase an exceptional capability between eukaryotic enzymes that is able to consume both NTPs and dNTPs (5, 11, 13). Remarkably, in primer synthesis, PrimPol essentially displays a predilection for dNTPs over NTPs a feature more characteristically allied with archaean primases (11).

PrimPol as a primase commotion is reliant on whole ZnF domain, which is dependable with prior studies on the herpes simplex virus type 1 (HSV1) helicase/primase complex. At this point, when fundamental scums in the UL52



zinc-binding domain were mutated primase activity was vanished (14, 15). Excitingly, zinc finger (ZnF) domain of PrimPol not to bind double-stranded (dsDNA) but single-stranded (ssDNA), proposing that for stabilizing PrimPol on ssDNA templates to allow synthesis of the initial dinucleotide this component may be significant (16). The capability of PrimPol to synthesis *de novo* DNA primers contributes it the prospective to reprime and resurrect replication downstream of DNA damage lesions and fork-stalling complications *in vivo* (17).

PrimePol Acts as a Polymerase

Additionally, DNA and RNA primase activity of PrimPol, it is also a template-dependent DNA polymerase, with capability to circumvent a numeral of DNA damage abrasions. Especially, PrimPol together oxidative and ultraviolet (UV)-induced lesions, as well as 8-oxo-guanine (8oxoG) and pyrimidine (6-4) pyrimidone photoproducts (6-4PPs) can circumvent (5, 11). Though, previous reports the exactness of circumvent varies, only 50% error-free in various occasions (11, 16, 18, 19). Previously, described effectiveness and mechanism of circumvent differs among reports, possibly brilliant metamorphoses in the metal cofactor used. Like, while manganese as a cofactor is cast-off, PrimPol can execute template reorganization to permit circumvent of cyclobutane pyrimidine dimers (CPDs), abasic sites (Ap sites) and 6-4 photoproducts (5, 20).

PrimPol in Mitochondrial DNA Replication

Genetic information majority is stored in the nucleus of mammalian cells. On the other hand, in the mitochondria a lesser amount of DNA is also found. Although mtDNA with (~16.6 kb) extended and encoding unbiased 13 polypeptides, (22 tRNAs and 2 rRNAs) but no introns mitochondrial genome mutation is liable for a numeral of mitochondriopathies and associated in several other pathologies with cardiovascular diseases, cancer and neurodegenerative complaints (21). Merely two non-coding regions exist (NCR), comprising the origin of DNA replication and the transcription origination start sites heavy-strand promoters (HSP), light-strand promoters (LSP) and O_L , (origin of light-strand DNA replication) respectively (7). Mitochondrial DNA (mtDNA) in mammalian cells is arranged into nucleoid assemblies having (~1-2) copies of the genome. While this is possible to differ provisionally nature of tissue and energy demand (22-24).

One salient feature of mammalian mitochondrial DNA (mtDNA) establishes inside the non-coding region (NCR) a little triple-stranded region. Where the count of a third DNA filament with 0.5 kb, named 7S DNA, forms a constant transposition D-loop structure (25). Electron micrograph images of mitochondrial DNA (mtDNA) were leading recognized the D-loop and later named due to its sedimentation rate in caesium chloride ascent studies the 7S DNA was originate (25, 26). In the mitochondrial genome wide-ranging protein are intricate in the replication, regulation and organization. Inside the nuclear genome all of these proteins are encoded and into the organelle must be transported as required (27, 28).

To confine to the mitochondria a substantial amount of PrimPol has been found. Where it relates with mitochondrial single-strand binding proteins (mtSSB), proposing a prospective part in the lenience of mitochondrial DNA (mtDNA) impairment (5, 11, 29, 30).

Self-Regulation of PrimPol

Traversed structural topographies to PrimPol by advantage of existence a primase also turn as essential regulatory

mechanisms. As revealed before, PrimPol shows very low processivity. Due to two significant features this particular description seems. First, the catalytic realm of archaeo-eukaryotic primase (AEP) than record polymerases has much reduced impression, elucidating conceivably why so out of sorts the enzyme dilemmas to DNA (31). Second, adversely settle acts of zinc finger (ZnF) realm to PrimPol's processivity (16). The regulatory consequence of the zinc finger (ZnF) domain, allows priming but confines elongation, joined with the archaeo-eukaryotic primase (AEP) reduced affection for DNA, limits the capability of PrimPol to contribute in substantial unfettered DNA synthesis through DNA replication (17).

CONCLUSIONS

Howard Flanders and Rupp acknowledge almost half a century before the existence of single stranded DNA (ssDNA) openings missing contrary UV photoproducts subsequent DNA replication in nucleotide elimination renovation lacking *E.coli* (32). Up to nearly four years the thought of leading strand re-instigation stayed provocative far ahead when origin-independent leading strand re-instigation was pragmatic (33). In the recent past, data have collected to provide leading strand re-priming as a well-maintained mechanism relative to replisome delaying challenges in eukaryotes (34-36). Latest investigations have proven that in eukaryotic organisms PrimPol's key role is to turn as a primase that helps an extensive array of leading strand impediments detour (16, 20, 37-40). The principal recommendation of re-priming, an exemplary was suggested which anticipated re-instigation of duplication downward of the impairment on both leading and lagging strand stencils.

Investigations from a numeral of test site have significantly amplified our appreciative of the role, recruitment and regulation of the enzyme during DNA replication (5, 11, 13). Though, we are simply just start to increase in value the versatile roles that play PrimPol in eukaryote.

Major role for PrimPol in DNA replication is emergent while leading strand re-priming, such as transcription the catalytic flexibility of the enzyme may provide itself to contrasting roles in further developments (41). It has been suggest that PrimPol is extremely acting the roles by re-priming DNA replication in mitochondria to permit an effective and appropriate way replication to be accomplished (7, 17).

PrimPol parameter seems a well streak amid inhibiting and affecting genetic flux, integrally PrimPol is more prone to error and also been in certain cancers such as glioma initiate to be over marked (30, 42).

Additional studies will be significant in PrimPol perspective defining the efficacy and capability of influencing in treating malignancy and unlike other ailments.

REFERENCES

1. Alberts B. The cell as a collection of protein machines: preparing the next generation of molecular biologists. *Cell*. 1998;92(3):291-4.
2. Hübscher U, Maga G, Spadari S. Eukaryotic DNA polymerases. *Annual review of biochemistry*. 2002;71(1):133-63.
3. Stucki M, Stagliar I, Jonsson ZO, Hübscher U. A coordinated interplay: proteins with multiple functions in DNA replication, DNA repair, cell cycle/checkpoint control, and transcription. 2000.
4. Goodman MF, Tiffin B. Sloppier copier DNA polymerases involved in genome repair. *Current opinion in genetics & development*. 2000;10(2):162-8.
5. García-Gómez S, Reyes A, Martínez-Jiménez MI, Chocrón ES, Morón S, Terrados G, et al. PrimPol, an archaic primase/polymerase



operating in human cells. *Molecular cell*. 2013;52(4):541-53.

6. Bolden A, Noy GP, Weissbach A. DNA polymerase of mitochondria is a gamma-polymerase. *Journal of Biological Chemistry*. 1977;252(10):3351-6.

7. Bailey LJ, Doherty AJ. Mitochondrial DNA replication: a PrimPol perspective. *Biochemical Society Transactions*. 2017;45(2):513-29.

8. Frick DN, Richardson CC. DNA primases. *Annual review of biochemistry*. 2001;70(1):39-80.

9. Aravind L, Leipe DD, Koonin EV. Toprim—a conserved catalytic domain in type IA and II topoisomerases, DnaG-type primases, OLD family nucleases and RecR proteins. *Nucleic acids research*. 1998;26(18):4205-13.

10. Iyer LM, Koonin EV, Leipe DD, Aravind L. Origin and evolution of the archaeo-eukaryotic primase superfamily and related palm-domain proteins: structural insights and new members. *Nucleic acids research*. 2005;33(12):3875-96.

11. Bianchi J, Rudd SG, Jozwiakowski SK, Bailey LJ, Soura V, Taylor E, et al. PrimPol bypasses UV photoproducts during eukaryotic chromosomal DNA replication. *Molecular cell*. 2013;52(4):566-73.

12. Rudd SG, Glover L, Jozwiakowski SK, Horn D, Doherty AJ. PPL2 translesion polymerase is essential for the completion of chromosomal DNA replication in the African trypanosome. *Molecular cell*. 2013;52(4):554-65.

13. Wan L, Lou J, Xia Y, Su B, Liu T, Cui J, et al. hPrimPol1/CCDC111 is a human DNA primase-polymerase required for the maintenance of genome integrity. *EMBO reports*. 2013;14(12):1104-12.

14. Biswas N, Weller SK. A mutation in the C-terminal putative Zn²⁺ finger motif of UL52 severely affects the biochemical activities of the HSV-1 helicase-primase subcomplex. *Journal of Biological Chemistry*. 1999;274(12):8068-76.

15. Chen Y, Carrington-Lawrence SD, Bai P, Weller SK. Mutations in the putative zinc-binding motif of UL52 demonstrate a complex interdependence between the UL5 and UL52 subunits of the human herpes simplex virus type 1 helicase/primase complex. *Journal of virology*. 2005;79(14):9088-96.

16. Keen BA, Jozwiakowski SK, Bailey LJ, Bianchi J, Doherty AJ. Molecular dissection of the domain architecture and catalytic activities of human PrimPol. *Nucleic acids research*. 2014;42(9):5830-45.

17. Guillian T, Doherty A. PrimPol—prime time to reprime. *Genes*. 2017;8(1):20.

18. Guillian TA, Bailey LJ, Brissett NC, Doherty AJ. PolDIP2 interacts with human PrimPol and enhances its DNA polymerase activities. *Nucleic acids research*. 2016;44(7):3317-29.

19. Stojković G, Makarova AV, Wanrooij PH, Forslund J, Burgers PM, Wanrooij S. Oxidative DNA damage stalls the human mitochondrial replisome. *Scientific reports*. 2016;6:28942.

20. Mourón S, Rodríguez-Acebes S, Martínez-Jiménez MI, García-Gómez S, Chocrón S, Blanco L, et al. Repriming of DNA synthesis at stalled replication forks by human PrimPol. *Nature structural & molecular biology*. 2013;20(12):1383.

21. Alexeyev M, Shokolenko I, Wilson G, LeDoux S. The maintenance of mitochondrial DNA integrity—critical analysis and update. *Cold Spring Harbor perspectives in biology*. 2013;5(5):a012641.

22. Iborra FJ, Kimura H, Cook PR. The functional organization of mitochondrial genomes in human cells. *BMC biology*. 2004;2(1):9.

23. Legros F, Malka F, Frachon P, Lombès A, Rojo M. Organization and dynamics of human mitochondrial DNA. *Journal of cell science*. 2004;117(13):2653-62.

24. Kukat C, Wurm CA, Spähr H, Falkenberg M, Larsson N-G, Jakobs S. Super-resolution microscopy reveals that mammalian mitochondrial nucleoids have a uniform size and frequently contain a single copy of mtDNA. *Proceedings of the National Academy of Sciences*. 2011;108(33):13534-9.

25. Kasamatsu H, Robberson DL, Vinograd J. A novel closed-circular mitochondrial DNA with properties of a replicating intermediate. *Proceedings of the National Academy of Sciences*. 1971;68(9):2252-7.

26. Walberg MW, Clayton DA. Sequence and properties of the human KB cell and mouse L cell D-loop regions of mitochondrial DNA. *Nucleic acids research*. 1981;9(20):5411-21.

27. Duxin JP, Dao B, Martinsson P, Rajala N, Guittat L, Campbell JL, et al. Human Dna2 is a nuclear and mitochondrial DNA maintenance protein. *Molecular and cellular biology*. 2009;29(15):4274-82.

28. Zheng L, Zhou M, Guo Z, Lu H, Qian L, Dai H, et al. Human DNA2 is a mitochondrial nuclease/helicase for efficient processing of DNA replication and repair intermediates. *Molecular cell*. 2008;32(3):325-36.

29. Bianchi J. Investigating the role of a novel primase-polymerase, PrimPol, in DNA damage tolerance in vertebrate cells: University of Sussex; 2013.

30. Guillian TA, Jozwiakowski SK, Ehlinger A, Barnes RP, Rudd SG, Bailey LJ, et al. Human PrimPol is a highly error-prone polymerase regulated by single-stranded DNA binding proteins. *Nucleic acids research*. 2014;43(2):1056-68.

31. Rechkoblit O, Gupta YK, Malik R, Rajashankar KR, Johnson RE, Prakash L, et al. Structure and mechanism of human PrimPol, a DNA polymerase with primase activity. *Science advances*. 2016;2(10):e1601317.

32. Rupp WD, Howard-Flanders P. Discontinuities in the DNA synthesized in an excision-defective strain of *Escherichia coli* following ultraviolet irradiation. *Journal of molecular biology*. 1968;31(2):291-304.

33. Heller RC, Marians KJ. Replication fork reactivation downstream of a blocked nascent leading strand. *Nature*. 2006;439(7076):557.

34. Lopes M, Foiani M, Sogo JM. Multiple mechanisms control chromosome integrity after replication fork uncoupling and restart at irreparable UV lesions. *Molecular cell*. 2006;21(1):15-27.

35. Elvers I, Johansson F, Groth P, Erixon K, Helleday T. UV stalled replication forks restart by re-priming in human fibroblasts. *Nucleic acids research*. 2011;39(16):7049-57.

36. Karras GI, Jentsch S. The RAD6 DNA damage tolerance pathway operates uncoupled from the replication fork and is functional beyond S phase. *Cell*. 2010;141(2):255-67.

37. Kobayashi K, Guillian TA, Tsuda M, Yamamoto J, Bailey LJ, Iwai S, et al. Repriming by PrimPol is critical for DNA replication restart downstream of lesions and chain-terminating nucleosides. *Cell Cycle*. 2016;15(15):1997-2008.

38. Schiavone D, Jozwiakowski SK, Romanello M, Guilbaud G, Guillian TA, Bailey LJ, et al. PrimPol is required for replicative tolerance of G quadruplexes in vertebrate cells. *Molecular cell*. 2016;61(1):161-9.

39. Pilzecker B, Buoninfante OA, Pritchard C, Blomberg OS, Huijbers IJ, van den Berk PC, et al. PrimPol prevents APOBEC/AID family mediated DNA mutagenesis. *Nucleic acids research*. 2016;44(10):4734-44.

40. Vallerga MB, Mansilla SF, Federico MB, Bertolin AP, Gottifredi V. Rad51 recombinase prevents Mre11 nuclease-dependent degradation and excessive PrimPol-mediated elongation of nascent DNA after UV irradiation. *Proceedings of the National Academy of Sciences*. 2015;112(48):E6624-E633.



41. Martínez-Jiménez MI, García-Gómez S, Bebenek K, Sastre-Moreno G, Calvo PA, Díaz-Talavera A, et al. Alternative solutions and new scenarios for translesion DNA synthesis by human PrimPol. DNA repair. 2015;29:127-38.
42. Yan X, Ma L, Yi D, Yoon J-g, Diercks A, Foltz G, et al. A CD133-related gene expression signature identifies an aggressive glioblastoma subtype with excessive mutations. Proceedings of the National Academy of Sciences. 2011;108(4):1591-6.

